### **Case Study / Discussion**

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### Scenario

- Drug "X": NCE / anti-epileptic
- No significant pre-clinical issues
- Phase I (6 studies)
  - significant PK variability
  - marked AEs of rash and / or cough (n=14/72)
- Phase II (initial POC study)
  - significant PK variability noted again
  - similar AE profile & incidence (n=8/35)

### **Metabolism Information**

- Preclinical in vitro studies show that Drug X is metabolized predominately by
  - CYP3A4 and CYP3A5 (accounts for >70%)
  - and to a lesser extent by CYP2C19 (<30%)
- In vitro data suggests that Drug X interacts with transporter genes ABCB1 and potentially ABCG2
- Note: CYP2C19 is a "known valid biomarker"

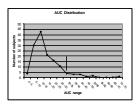
### Inter-individual variation of Drug X plasma concentrations

Inter-individual variation of Drug X plasma concentrations has been consistently observed in both Phase I & Phase II

- AUC range 1.43 73.23 ng.hr/ml
- Cmax range 0.258 4.511 ng/ml
- Tmax range 0.75 12 hours
- 10-20% incidence of "outliers" with marked PK variability
- Note: Not all subjects with marked PK variability had rash and/or cough (AE incidence approx. 20%)

### Pharmacogenetics and PK / AE - analyses

- Evaluation (retrospective) of Drug X examining association of pharmacokinetic (PK) variability, rash and cough (18 subjects with defined moderate/severe AEs)
- Combined Ph I & Ph II studies (7 in total) to increase power
- Five candidate genes known to play a role in metabolism of Drug X: CYP2C19, CYP3A4, CYP3A5 and ABCB1, ABCG2
- Subjects: 107 subjects in total 72 healthy volunteers from six phase I studies and 35 patients from a Ph II study



### Subject/Phenotype Breakdown

Phenotype	# of 'phenotypic events' (ph I / Ph II)	# of 'controls'
AUC	13 ( 9/4)	87
Cmax	10 ( 7/3)	90
Tmax	19 ( 13/6)	81
Cough	18 ( 11/7)	89
Rash	15 ( 10/5)	92

## Analyses Conducted Retrospectively after Ph I studies and initial Ph II (7 studies combined)

- Single-point (genotypic and allelic) association analyses of SNPs
- Hardy-Weinberg analysis to confirm genetic segregation
- Linkage disequilibrium (LD) analyses

#### **CYP2C19**

- 22 SNPs within the CYP2C19 gene showed association (p<0.01) with incidence of **rash** and 6 of these SNPs showed association (p<0.01) with incidence of **cough**
- CYP2C19 \*2/\*2 genotypic p -value was p=0.001 for cough and p=0.0016 for rash
- LD in CYP2C19 was generally low; not all of the SNPs associated with rash and cough were in strong LD
- Additionally, 13 different SNPs in CYP2C19 showed association with Tmax (p<0.01)</li>

### ABCB1

- 16 SNPs in the ABCB1 gene showed single-point association with Tmax (p<0.01) in Caucasians</li>
- the 16 SNPs noted above were not in strong LD
- 4 SNPs in ABCB1 showed association with AUC and Cmax

### **Summary of Significant Results**

- Association was observed between SNPs in CYP2C19 with rash, cough and Tmax
- Association was observed between SNPs in ABCB1 with Tmax
- No evidence for association was observed in ABCG2, CYP3A4 or CYP3A5
- 3 of 3 subjects homozygous for CYP2C19\*2 had rash and cough

# DISCUSSION AUDIENCE AND PANEL PARTICIPATION

Firstly, let's consider what might be done differently vs what the Sponsor did:

- (i) Prior to going into FTIH, would you have planned to proactively genotype in Phase I?
- No
- 2. Yes (CYP3A4, CYP3A5, CYP2C19, ABCB1, ABCG2)
- 3. CYP3A4, CYP3A5 and CYP2C19 only
- 4. A more comprehensive DME panel

<ul> <li>(ii) Following review of the phenotypic (PK &amp; AE) data from the 6 completed Phase I studies, would you have retrospectively genotyped samples from those studies at that point?</li> <li>1. No</li> <li>2. Yes (CYP3A4, CYP3A5, CYP2C19, ABCB1, ABCG2)</li> <li>3. CYP3A4, CYP3A5 and CYP2C19 only</li> <li>4. A more comprehensive DME panel</li> </ul>	(iii) Would your response to the previous question be affected if the major metabolic pathway was via a "known valid biomarker"?  1. Yes 2. No
<ul> <li>(iv) Following review of the phenotypic (PK &amp; AE) data from the 6 completed Ph I studies, would you have planned to proactively genotype samples from the next Phase II POC study?</li> <li>1. No</li> <li>2. Yes (CYP3A4, CYP3A5, CYP2C19, ABCB1, ABCG2)</li> <li>3. CYP3A4, CYP3A5 and CYP2C19 only</li> <li>4. A more comprehensive DME panel</li> </ul>	(v) Would your response to the previous question be affected if the major metabolic pathway was via a "known valid biomarker"?  1. Yes 2. No
Now let's turn to the results that the Sponsor actually obtained:  (vi) Would you prospectively plan to replicate the findings / confirm the data (e.g. in a follow up Ph II study)?  1. Yes 2. No	(vii) If the initial data are confirmed / replicated, would you prospectively stratify in a subsequent study for dosing (e.g. Ph III)?  1. Yes 2. No

(viii) If the initial data are confirmed/replicated, do you consider that the genetic association data sufficiently demonstrate a correlation between PK variability and the SNPs analysed such that continued evaluations be carried out?  1. Yes 2. No	<ul> <li>(ix) If the initial data are confirmed/replicated, do you consider that the genetic association data sufficiently demonstrate a correlation between AEs and the SNPs analysed such that continued evaluations be carried out?</li> <li>1. Yes</li> <li>2. No</li> </ul>
<ul> <li>(x) If the initial data are confirmed/replicated, do you consider that this may constitute the basis for pursuing development of Drug X with a diagnostic test for potential prediction of optimal dosing?</li> <li>1. Yes</li> <li>2. No</li> </ul>	<ul> <li>(xi) If the initial data are confirmed/replicated, do you consider that this may constitute the basis for pursuing development of Drug X with a diagnostic test for potential prediction of AEs (rash, cough)?</li> <li>1. Yes</li> <li>2. No</li> </ul>
<ul> <li>(xii) If the initial data are confirmed/replicated, do you consider that these data may have implications for identifying those patients most likely to show efficacy?</li> <li>1. Yes</li> <li>2. No</li> </ul>	Acknowledgements